

STRUCTURE AND STEREOCHEMISTRY OF EUPAFORMONIN, A NOVEL
CYTOTOXIC SESQUITERPENE LACTONE FROM *EUPATORIUM FORMOSANUM* HAY.

Andrew T. McPhail* and Kay D. Onan

Paul M. Gross Chemical Laboratory, Duke University,
Durham, North Carolina 27706

Kuo-Hsiung Lee* and Toshiro Ibuka

Department of Medicinal Chemistry, School of Pharmacy,
University of North Carolina, Chapel Hill, North Carolina 27514

Huan-Chang Huang

School of Pharmacy, Kaohsiung Medical College,
Kaohsiung, Taiwan

(Received in USA 24 May 1974; received in UK for publication 26 July 1974)

The whole plant of *Eupatorium formosanum* HAY. collected in the early spring in Tainan, Taiwan, was previously reported to yield eupatolide¹ (I) (0.4%) as the major cytotoxic agent. A new collection of the plant from a different location² has led to the isolation of a new cytotoxic³ sesquiterpene lactone, eupafomonin (II) (0.002%), in addition to eupatolide (0.001%).

Eupafomonin was isolated as colorless prisms from the ethanolic extract of *E. formosanum* according to an exact literature procedure¹ followed by silica gel column chromatography⁵. Eupafomonin [II, C₁₇H₂₂O₅, m.p. 216-218°, m/e 306.1461 (M⁺), 264 (M-42) (M-COCH₃), 246 (M-60) (M-CH₃COOH), and 228 (M-60-18) (M-CH₃COOH-H₂O); ν_{\max} (Nujol) 3409 (OH), 1753 (γ -lactone), 1712 (C=O), 1675 and 1663 cm⁻¹ (C=C); δ (pyridine-d₅) 6.41 (1H, d, J = 3.0 Hz, 13-H), 5.72 (1H, d, J = 3.0 Hz, 13H), 2.00 (3H, s, OCOCH₃), 2.12 (3H, s) and 1.79 (3H, s) (two vinyl methyl protons)] gave, upon treatment with acetic anhydride in pyridine, an acetate (III) in 50% yield⁶. Compound (III) (C₁₉H₂₄O₆, m.p. 178°, colorless needles) showed ν_{\max} (CHCl₃) 1759 (γ -lactone), 1742 (ester C=O) and 1662 cm⁻¹ (C=C); and δ (CDCl₃) 2.12 (3H, s), 2.06 (3H, s) (two OCOCH₃), 1.92 (3H, s) and 1.79 (3H, s) (two vinyl methyl groups).

The complete molecular structure and relative stereochemistry of eupafomonin were established by a single crystal X-ray analysis. Eupafomonin crystallizes in the orthorhombic system, space group *P*2₁2₁2₁, *a* = 14.28(1), *b* = 11.56(1), *c* = 9.68(1) Å, *Z* = 4. The crystal

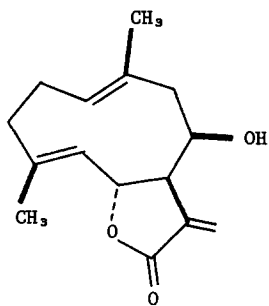
structure was elucidated by direct phase determining methods using MULTAN⁷ and refined by full-matrix least-squares calculations to the present $R = 0.106$ over 1437 visually estimated reflections from photographic data taken with $\text{Cu-K}\alpha$ ($\lambda = 1.542 \text{ \AA}$) radiation.

The X-ray study proves that eupafornonin has constitution (II) and if we assume that the C₇ hydrogen atom is α -oriented as in all known naturally-occurring germacranolides, then (II) also represents the absolute configuration. Eupaformonin is therefore a new member of the $\Delta^{1(10)}$ *trans*, $\Delta^{4(5)}$ *cis* germacradiene class of sesquiterpenes, sub-group 'heliangolides', all previous authentic examples⁸ of which have been derivatives of helianginol (IV) and generally have a 3β -hydroxyl function in addition to the 1(10)-epoxide group; typical examples are heliangine^{14,15} (V), erioflorin¹⁶ (VI), and erioflorin methacrylate¹⁷ (VII). On the basis of these consistent, but limited, observations it was speculated¹⁷ that the $\Delta^{4(5)}$ *cis* bond may arise from a *trans*-oriented precursor after (or during) the introduction of the oxygen functions. The existence of eupafornonin with its 3α -acetate group and 1(10) *trans* double bond now indicates that the biosynthetic stage at which the *trans-cis*-isomerization occurs may equally well precede the oxidation step.

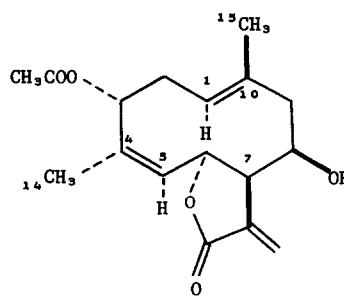
In (II) the cyclodecadiene ring adopts the boat-chair conformation (VIII) with the C₁₄ and C₁, methyl substituents *anti* with respect to the ring plane. Endocyclic torsion angles around the *cis* and *trans* double bonds are 3° and 169° , respectively. Whereas the former is not significantly different from the ideal unstrained 0° value, the torsion angle at the *trans* double bond departs significantly from 180° and reflects the more strained nature of this bond. The angle found here is similar in magnitude to the corresponding values at the *trans* double bonds in shiromodiol¹⁸ (167°), elephantol¹⁹ (163°), and dihydromikanolide²⁰ (-163°) but somewhat less than that in the more severely strained melampodin²¹ (-155°).

Acknowledgement

We thank Dr. David L. Harris, Department of Chemistry, University of North Carolina at Chapel Hill for XL-100 n.m.r. spectra, and Dr. David Rosenthal and Mr. Fred Williams of the Research Triangle Center for Mass Spectrometry for mass spectral data. This investigation was supported in part by U. S. Public Health Service Research Grant No. CA 12360 from the National Cancer Institute to K.H.L.

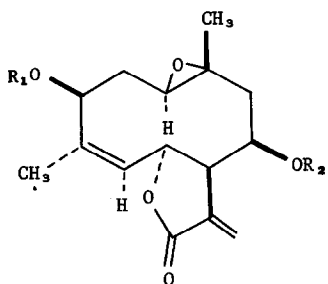


(I)

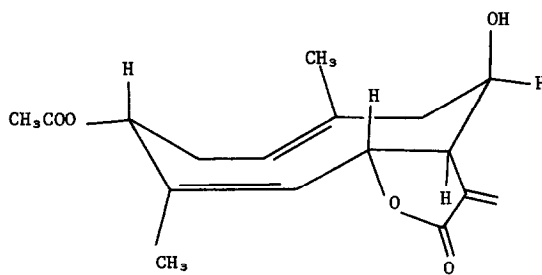
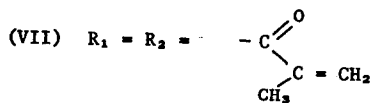
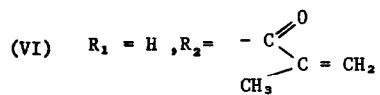
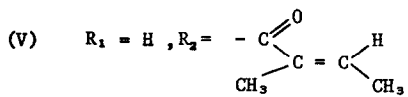


(II) R = H

(III) R = COCH₃



(IV) R₁ = R₂ = H



(VIII)

References and Footnotes

1. K. H. Lee, H. C. Huang, E. S. Huang, and H. Furukawa, J. Pharm. Sci., 61, 629 (1972).
2. Specimens were gathered in early spring, 1973, in Wootai, Pingtung, Taiwan.
3. Eupaformonin showed significant inhibitory activity of the *in vitro* growth of tissue culture cells originating from human epidermoid carcinoma of larynx (H. Ep.-2). Cytotoxicity was assayed by Dr. E. S. Huang, Department of Bacteriology and Immunology School of Medicine, University of North Carolina at Chapel Hill, by literature method⁴.
4. E. S. Huang, K. H. Lee, C. Piantadosi, T. A. Geissman, and J. S. Pagano, J. Pharm. Sci., 61, 1960 (1972).
5. Mallinckrodt silica gel CC7 was used, and the column was eluted with chloroform-ethyl acetate (9:1).
6. Starting material was recovered in 50% yield. The axial orientation of the C₈ hydroxyl group as shown by the X-ray analysis may well account for the difficulty of this acetylation.
7. G. Germain, P. Main, and M. M. Woolfson, Acta Cryst., A27, 368 (1971).
8. Classification of several other $\Delta^{4(s)}$ *cis* germacranolide sesquiterpene lactones into the recently proposed sub-groups^{9,10} cannot be made unambiguously for the *cis*, *trans* nature of the normal reference 1(10) double bond or its equivalent is obscured e.g. eupacunin¹¹, liatrin¹², or woodhousin¹³.
9. S. Neidle and D. Rogers, J.C.S. Chem. Comm., 140 (1972).
10. D. Rogers, G. P. Moss, and S. Neidle, J.C.S. Chem. Comm., 142 (1972).
11. S. M. Kupchan, M. Maruyama, R. J. Hemingway, J. C. Hemingway, S. Shibuya, T. Fujita, P. D. Cradwick, A. D. U. Hardy, and G. A. Sim, J. Amer. Chem. Soc., 93, 4914 (1971); S. M. Kupchan, M. Maruyama, R. J. Hemingway, J. C. Hemingway, S. Shibuya, and T. Fujita, J. Org. Chem., 38, 2189 (1973).
12. S. M. Kupchan, V. H. Davies, T. Fujita, M. R. Cox, and R. F. Bryan, J. Amer. Chem. Soc., 93, 4916 (1971); S. M. Kupchan, V. H. Davies, T. Fujita, M. R. Cox, R. J. Restivo, and R. F. Bryan, J. Org. Chem., 38, 1853 (1973).
13. W. Herz and S. V. Bhat, J. Org. Chem., 37, 906 (1972).
14. N. Nishikawa, K. Kamiya, A. Takabatake, H. Oshio, Y. Tomie, and I. Nitta, Tetrahedron, 22, 3601 (1966).
15. It has been pointed out elsewhere⁹ that in the original publication¹⁴ the constitutional formula for heliangine was drawn incorrectly as it showed $\Delta^{4(s)}$ *trans* although the crystallographic co-ordinates and molecular diagram indicate a $\Delta^{4(s)}$ *cis* double bond. Unfortunately, while ref. 9 correctly showed the double bond configuration, C₃ and its hydroxyl substituent were omitted. Formula (V) here correctly represents heliangine and erioflorin (VI) which was based upon it.¹⁶
16. S. J. Torrance, T. A. Geissman, and M. R. Chedekel, Phytochemistry, 8, 2381 (1969).
17. S. Gnecco, J. P. Poyser, M. Silva, P. G. Sammes, and T. W. Tyler, Phytochemistry, 12, 2469 (1973).
18. R. J. McClure, G. A. Sim, P. Coggon, and A. T. McPhail, Chem. Comm., 128 (1970).
19. A. T. McPhail and G. A. Sim, J. Chem. Soc. Perkin II, 1313 (1972).
20. P. J. Cox, G. A. Sim, J. S. Roberts, and W. Herz, J.C.S. Chem. Comm., 428 (1973).
21. S. F. Watkins, N. H. Fischer, and I. Bernal, Proc. Nat. Acad. Sci., 70, 2434 (1973).